

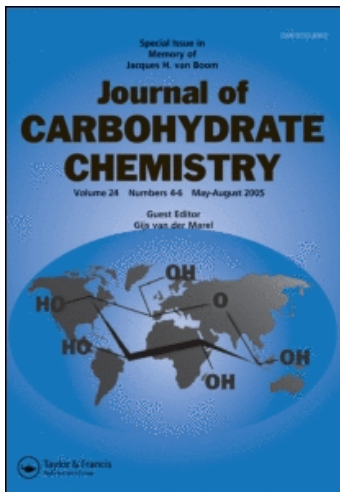
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Synthesis of the Dimethyl Ester of 1-Deoxy-1-Idonojirimycin-1-Methylenphosphonate: A New Approach to Iminosugar Phosphonates

Barbara La Ferla^a; Piergiuliano Bugada^a; Francesco Nicotra^a

^a Department of Biotechnology and Bioscience, University of Milano Bicocca, Milano, Italy

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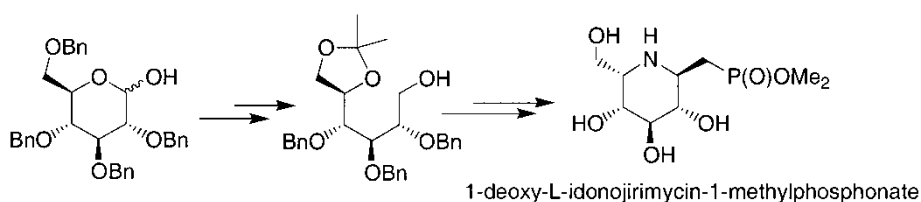
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Synthesis of the Dimethyl Ester of 1-Deoxy-L-Idonojirimycin-1-Methylenphosphonate: A New Approach to Iminosugar Phosphonates

Barbara La Ferla, Piergiuliano Bugada, and Francesco Nicotra

Department of Biotechnology and Bioscience, University of Milano Bicocca, Milano, Italy

1-Methylenphosphonate-1-deoxy-L-idonojirimycin (**1**) has been synthesized starting from commercially available tetrabenzyl glucose, the key steps being substitution of the hydroxyl group at C-5 of compound **7** with an azido group, stereoselective reaction of the aldehyde at C-1 of compound **10** with dimethyl methylenephosphonate anion, conversion of the azide into an amino group, and finally cyclization of the aminoalcohol **12**.



Keywords Carbohydrate analogs, Iminosugars, Phosphonates, Inhibitors

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Address correspondence to Barbara La Ferla, Department of Biotechnology and Bioscience, University of Milano Bicocca, Piazza della Scienza 2, I-20126 Milano, Italy.
E-mail: barbara.laferla@unimib.it

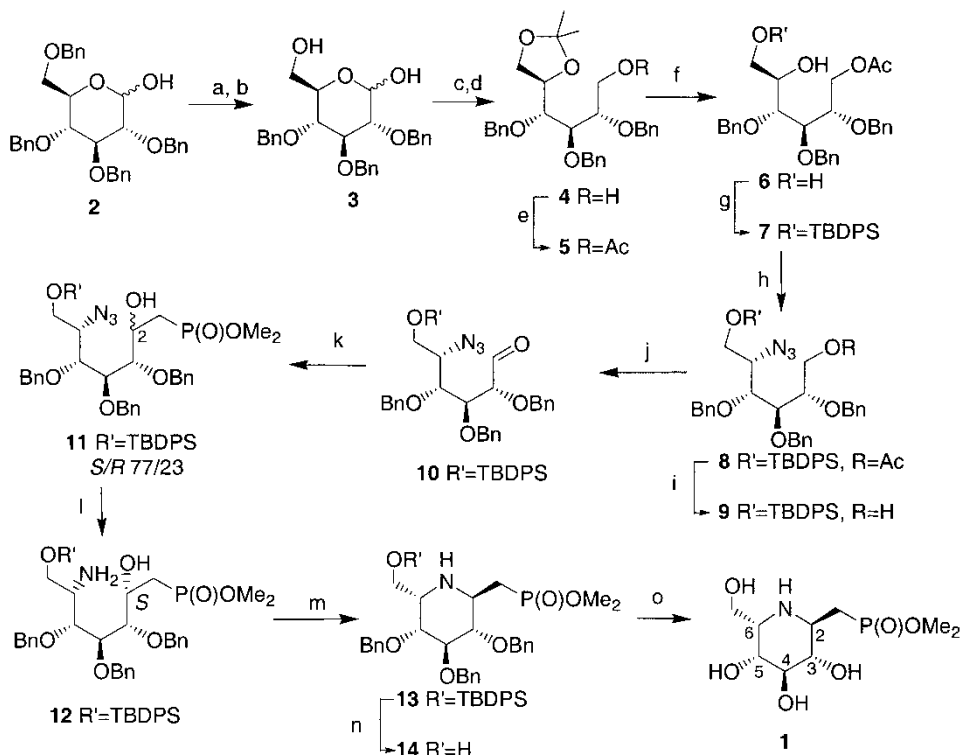
INTRODUCTION

Naturally occurring iminosugars are known to be potent inhibitors of carbohydrate processing enzymes such as glycosidases and glycosyltransferases,^[1] which are involved in many biological and pathological processes. This has suggested their use in a wide range of potential therapeutic applications,^[2–6] and has stimulated synthetic chemists to develop new and more efficient synthetic strategies for the preparation of new, more potent and selective iminosugar derivatives. Due to the great interest in these compounds, a great number of novel structures have been synthesized so far, nonetheless, there are only very few examples of iminosugar phosphonates.^[7] In this work we report the synthesis of a new compound 1-deoxy-L-idonojirimycin-1-methylenephosphonate using a novel approach to iminosugar phosphonates.

RESULTS AND DISCUSSION

Our synthetic route started from tetrabenzyl glucose **2**. In order to introduce a nitrogen atom at C-5, which was required for the generation of the iminosugars, the following synthetic scheme was elaborated (Sch. 1): acetolysis and basic hydrolysis of the acetate afforded compound **3**, which in turn was reduced to the corresponding alditol, which was protected as isopropylidene derivative **4**. Protection of the primary hydroxyl group at C-1 as acetate (**5**) and cleavage of the isopropylidene group followed by protection of the primary hydroxyl group at C-6 as *t*-butyldiphenylsilyl ether afforded compound **7** with a free hydroxyl group at C-5. The free hydroxyl group of **7** was exploited for the introduction of the nitrogen atom; its substitution with an azide through a Mitsunobu reaction afforded compound **8** with inversion of configuration at C-5. In order to introduce the methylenephosphonate moiety, the acetylated primary hydroxyl group at C-1 was selectively deprotected and oxidized to the corresponding aldehyde **10**, and then subjected to a nucleophilic attack using lithium methylenedimethyl phosphonate. The reaction afforded compound **11** as an inseparable mixture of diastereoisomers in a ratio of *R/S* = 23:77, determined by ¹H NMR spectroscopy. Only after reduction with triphenylphosphine in water/THF the corresponding major amine **12** was obtained as pure product in 51% yield over the two steps.

The absolute configuration of the newly formed stereocenter was attributed to the cyclized product and is discussed later on. Compound **12** was then cyclized through an intramolecular Mitsunobu reaction, affording, after removal of the silyl protection, pure compound **14** in 58% yield over the two steps. Final debenylation afforded the dimethyl ester of 1-deoxy-L-idonojirimycin-1-methylenephosphonate (**1**). ¹H NMR spectroscopic analysis of compound **14** allowed us to determine the absolute configuration of the C(2) stereocenter formed during the addition of the phosphonate group. The



Scheme 1: Reagents and conditions. a) $\text{Ac}_2\text{O}/\text{TFA}$ 4/1 then NaOH 4M; b) Na , MeOH ; c) NaBH_4 , EtOH ; d) 2,2-dimethoxypropane, camphor-10-sulfonic acid, CH_3CN (91% over four steps); e) Ac_2O , pyridine, DMAP, CH_2Cl_2 (96%); f) H_2O , CSA, CH_3CN , 60°C (96%); g) TBDPSCI, imidazole, CH_2Cl_2 (95%); h) $(\text{PhO})_2\text{PON}_3$, PPh_3 , DIAD, THF (71%); i) MeONa , MeOH (91%); j) Dess-Martin periodinane, CH_2Cl_2 ; k) $\text{CH}_3\text{PO}(\text{OMe})_2$, BuLi , THF, -78°C (62% over two steps); l) PPh_3 , H_2O , THF, 60°C (51%); m) PPh_3 , DIAD, THF; n) TBAF, THF (58% over two steps); o) $\text{Pd}(\text{OH})_2/\text{C}$, H_2 , MeOH/AcOH 98%.

values of the coupling constants ($J_{3,2} = 8.5$ Hz, $J_{4,3} = 8.0$ Hz, and $J_{5,4} = 9.0$ Hz) (Fig. 1) were indicative of a *trans*-diaxial disposition of the protons, thus indicating a 4C_1 conformation; moreover, the diaxial disposition of C(2)-H/C(3)-H allowed to determine the absolute configuration (*R*) of C(2).

Biological evaluation of compound 1 in a variety of glycosidase inhibition assay is in due course.

EXPERIMENTAL

General Remarks

All solvents were dried with molecular sieves for at least 24 h prior to use. Thin layer chromatography (TLC) was performed on silica gel 60 F254 plates

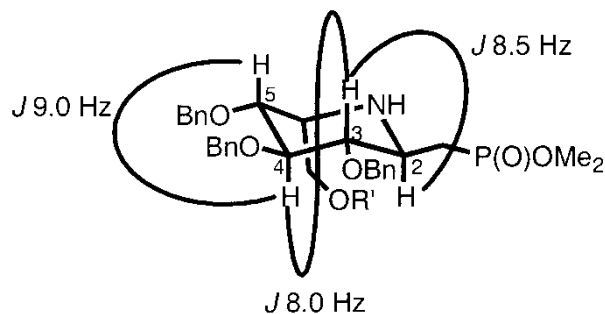


Figure 1: Conformation and coupling constants values of compound **13**.

(Merck) with detection using UV light when possible, or by charring with a solution of concd. $\text{H}_2\text{SO}_4/\text{EtOH}/\text{H}_2\text{O}$ (5:45:45) or a solution of $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}$ (21 g), $\text{Ce}(\text{SO}_4)_2$ (1 g), concd. H_2SO_4 (31 mL) in water (500 mL). Flash column chromatography was performed on silica gel 230–400 mesh (Merck). ^1H , ^{13}C NMR, and ^{31}P spectra were recorded at 25°C with a Varian Mercury 400 MHz instrument using CDCl_3 as the solvent unless otherwise stated. Chemical shift assignments, reported in ppm, are referenced to the corresponding solvent peaks. Mass spectra were recorded with a MALDI2 Kompakt Kratos instrument, using gentisic acid (DHB) as the matrix. Optical rotations were measured at rt using a Krüss P3002 electronic polarimeter and are reported in units of $10^{-1}\text{deg}\cdot\text{cm}^2\cdot\text{g}^{-1}$. Elemental analyses were performed using a Perkin-Elmer Series II Analyzer 2400.

2,3,4-Tri-O-benzyl-D-glucopyranose (3). Compound **2** (2.5 g, 4.62 mmol), was cooled to 0°C under an inert atmosphere, and a solution of $\text{Ac}_2\text{O}/\text{TFA}$ 4/1 (30 mL) was slowly added. The reaction was left stirring at 0°C . After 3 h ice-cold water was added (50 mL), and after 15 min the reaction mixture was neutralized by a slow addition at 0°C of NaOH 4M. The mixture was then extracted with AcOEt ($3 \times 100\text{ mL}$), the combined organic layers were dried over Na_2SO_4 , and the solvent was eliminated under reduced pressure. The crude was dissolved in dry MeOH (10 mL) and a catalytic amount of Na was added. After 15 min the reaction mixture was neutralized with amberlite IR 120- H^+ , filtered, and evaporated, affording known compound **3**^[8] as mixture of diastereoisomers. Crude **3** was directly used for the following step.

2,3,4-Tri-O-benzyl-5,6-O-isopropylidene-D-glucitol (4). Crude compound **3** was dissolved in EtOH (30 mL) and NaBH_4 (1.17 g, 31 mmol) was added in five portions. After 3 h the solvent was removed under reduced pressure and the solid residue was suspended in a saturated solution of Na_2CO_3 (30 mL). The suspension was stirred for 20 min and then extracted with EtOAc

(3 × 30 mL), the combined organic layers were dried over Na₂SO₄, and the solvent was eliminated under reduced pressure. The crude was dissolved in dry CH₃CN (10 mL) under inert atmosphere, and 2,2-dimethoxypropane (1.2 mL, 10.07 mmol) and a catalytic amount of camphor-10-sulfonic acid were added. After 1 h the reaction was neutralized with triethylamine and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (petroleum ether/ethyl acetate v/v 7/3), yielding **4** (3.48 g, 91% from **2**) as a colorless oil. $[\alpha]_D^{20} +1.2$ (c 0.4 CHCl₃); ¹H NMR: δ = 1.33 (s, 3 H, CH₃iPr), 1.42 (s, 3 H, CH₃iPr), 3.50 [m, 1 H, C(1a)-H], 3.70–3.78 [m, 3 H, C(2)-H, C(3)-H, C(1b)-H], 3.93–3.97 [m, 2 H, C(4)-H, C(6a)-H], 4.03 [dd, 1 H, *J* = 8.0 Hz, *J* = 7.2 Hz, C(6b)-H], 4.22 [bdt, 1 H, *J* = 7.2 Hz, *J* = 4.0 Hz, C(5)-H], 4.61–4.68 (m, 4 H, 4CHPh), 4.75 (d, 1 H, *J* = 11.2 Hz, CHPh), 4.83 (d, 1 H, *J* = 11.6 Hz, CHPh), 7.19–7.34 (m, 15 H, CHAR) ppm. ¹³C NMR: δ = 25.38, 26.97 (2 CH₃iPr), 61.87, 66.20 [C(1), C(6)], 73.30, 74.33, 75.05 (3CH₂Ph), 77.44, 78.33, 78.83, 79.88 [C(2), C(3), C(4), C(5)], 108.5 (CqiPr), 127.9–128.7 (CHAR), 138.1, 138.2, 138.3 (CqAr) ppm. MS (MALDI-TOF): *m/z* 515 [M + Na]⁺, 531 [M + K]⁺; C₃₀H₃₆O₆ (492.25): Calcd.; C, 73.15; H, 7.37. found; C, 72.95; H, 7.21.

1-O-Acetyl-2,3,4-tri-O-benzyl-5,6-O-isopropylidene-D-glucitol (5).

Compound **4** (183 mg, 0.37 mmol) was dissolved in CH₂Cl₂ (2 mL) and pyridine (6 equiv., 2.22 mmol, 176 mg, 179 μL), DMAP (0.1 equiv., 0.04 mmol, 5 mg), and acetic anhydride (3 equiv., 1.11 mmol, 114 mg, 105 μL) were added. The solution was stirred at rt for 1 h, and then HCl (5%, 2 mL) was added. The two layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 5 mL). The combined organic layers were dried over Na₂SO₄. The solvent was eliminated under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ethyl acetate v/v 10/1), yielding **5** (189 mg, 96%) as colorless oil. $[\alpha]_D^{20} +7.0$ (c 1.0 CHCl₃); ¹H NMR: δ = 1.23 (s, 3 H, CH₃iPr), 1.33 (s, 3 H, CH₃iPr), 1.93 (s, 3H, CH₃Ac), 3.58 [dd, 1 H, *J* = 6.1 Hz, *J* = 4.1 Hz, C(4)-H], 3.79 [ddd, 1 H, *J* = 9.4 Hz, *J* = 5.9 Hz, *J* = 3.3 Hz, C(2)-H], 3.83–3.87 [m, 2 H, C(3)-H, C(6a)-H], 3.92 [bt, 1 H, *J* = 7.6 Hz, C(6b)-H], 4.00 [dd, 1 H, *J* = 12.0 Hz, *J* = 5.9 Hz, C(1a)-H], 4.12 [ddd, 1 H, *J* = 11.3 Hz, *J* = 6.8 Hz, *J* = 4.2 Hz, C(5)-H], 4.25 [dd, 1 H, *J* = 12.0 Hz, *J* = 3.4 Hz, C(1b)-H], 4.51 (d, 1 H, *J* = 11.6 Hz, CHPh), 4.55 (d, 1 H, *J* = 11.3 Hz, CHPh), 4.57 (d, 1 H, *J* = 11.3 Hz, CHPh), 4.58 (d, 1 H, *J* = 11.6 Hz, CHPh), 4.64 (d, 1 H, *J* = 11.3 Hz, CHPh), 4.73 (d, 1 H, *J* = 11.3 Hz, CHPh), 7.19–7.24 (m, 15 H, CHAR) ppm. ¹³C NMR: δ = 21.43(CH₃Ac), 25.43, 26.97 (2 CH₃iPr), 64.23, 66.21 [C(1), C(6)], 73.35, 74.47, 74.95 (3CH₂Ph), 77.36, 77.54, 78.53, 79.33 [C(2), C(3), C(4), C(5)], 108.5 (CqiPr), 127.9–132.4 (CHAR), 138.2, 138.4, 138.4 (CqAr), 170.9 (C=O) ppm. MS (MALDI-TOF): *m/z* 557 [M + Na]⁺, 573 [M + K]⁺; C₃₂H₃₈O₇ (534.26): Calcd; C, 71.89; H, 7.16. Found; C, 71.95; H 7.20.

1-O-Acetyl-2,3,4-tri-O-benzyl-D-glucitol (6). Compound **5** (1.07 g, 2.01 mmol) was dissolved in CH₃CN (10 mL), and then H₂O (500 μL) and CSA (0.1 equiv., 0.2 mmol, 50 mg) were added. The mixture was stirred at 60°C for 20 min, and then NaHCO₃ (sat. sol., 5 mL) was added. The two layers were separated, and the aqueous layer was extracted with ethyl acetate (3 × 5 mL). The combined organic layers were dried over Na₂SO₄. The solvent was eliminated under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ethyl acetate v/v 8/2), yielding **6** (944 mg, 96%) as colorless oil. $[\alpha]_{\text{D}}^{20} +11.2$ (c 0.9 CHCl₃); ¹H NMR: δ = 1.89 (s, 3H, CH₃Ac), 2.22–2.28 (m, 1H, OH), 3.27 (d, 1H, *J* = 5.6 Hz, OH), 3.55–3.58 [m, 1H, C(6a)-H], 3.64–3.67 [m, 3H, C(3)-H, C(4)-H, C(6b)-H], 3.73–3.75 [m, 1H, C(5)-H], 3.84–3.87 [m, 1H, C(2)-H], 4.11 [dd, 1H, *J* = 11.5 Hz, *J* = 5.9 Hz, C(1a)-H], 4.19 [dd, 1H, *J* = 11.5 Hz, *J* = 4.8 Hz, C(1b)-H], 4.51–4.61 (m, 6H, 6CHPh), 7.15–7.26 (m, 15H, CHAr) ppm. ¹³C-NMR: δ = 21.37 (CH₃Ac), 63.78, 63.89 [C(1), C(6)], 72.06 [C(5)], 73.61, 74.01, 74.17 (3CH₂Ph), 76.62, 76.88, 78.20 [C(2), C(3), C(4)], 128.2–128.7 (CHAr), 137.6, 137.7, 137.9 (3CqAr), 170.8 (C=O) ppm. MS (MALDI-TOF): *m/z* 518 [M + Na]⁺, 534 [M + K]⁺; C₂₉H₃₄O₇ (494.23): Calcd: C, 70.43; H, 6.93. Found: C, 70.29; H, 7.03.

1-O-Acetyl-2,3,4-tri-O-benzyl-6-O-*t*-butyldiphenylsilyl-D-glucitol (7). Compound **6** (944 mg, 1.93 mmol) was dissolved, under inert atmosphere, in dry CH₂Cl₂ (10 mL), and imidazole (3 equiv., 5.79 mmol, 394 mg) and *tert*-butyldiphenylchlorosilane (1.5 equiv., 2.90 mmol, 741 μL) were added. The solution was stirred at rt overnight, and then CH₃OH (1 mL) and H₂O (10 mL) were added. The two layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic layers were dried over Na₂SO₄. The solvent was eliminated under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ethyl acetate v/v 9/1), yielding **7** (1.35 g, 95%) as colorless oil. $[\alpha]_{\text{D}}^{20} -1.7$ (c 1.4 CHCl₃); ¹H NMR: δ = 1.09 (s, 9H, CH₃*t*Bu), 1.98 (s, 3H, CH₃Ac), 2.85 (d, 1H, *J* = 5.2 Hz, OH), 3.82–3.88 [m, 4H, C(3)-H, C(4)-H, C(6a)-H, C(6b)-H], 3.90–3.98 [m, 2H, C(2)-H, C(5)-H], 4.17 [dd, 1H, *J* = 11.9 Hz, *J* = 6.1 Hz, C(1a)-H], 4.35 [dd, 1H, *J* = 11.8 Hz, *J* = 3.4 Hz, C(1b)-H], 4.55 (s, 2H, CH₂Ph), 4.61 (d, 1H, *J* = 11.4 Hz, CHPh), 4.64 (d, 1H, *J* = 10.9 Hz, CHPh), 4.67 (d, 1H, *J* = 10.9 Hz, CHPh), 4.69 (d, 1H, *J* = 11.4 Hz, CHPh), 7.24–7.67 (m, 25H, CHAr) ppm. ¹³C-NMR: δ = 19.75 (Cq*t*Bu), 21.39 (CH₃Ac), 27.36 (CH₃*t*Bu), 64.53, 65.19 [C(1), C(6)], 72.05 [C(5)], 73.50, 73.54, 74.72 (3CH₂Ph), 77.33, 77.48, 78.52 [C(2), C(3), (4)], 127.8–130.0 (CHAr), 133.2, 133.3 (2CqAr), 135.7, 135.8 (CHAr), 138.1, 138.2, 138.2 (3CqAr), 170.8 (C=O) ppm. MS (MALDI-TOF): *m/z* 756 [M + Na]⁺, 772 [M + K]⁺; C₄₅H₅₂O₇Si (732.35): calcd: C, 73.74; H, 7.15. Found: C, 73.94; H, 7.22.

1-O-Acetyl-5-azido-2,3,4-tri-O-benzyl-6-O-*t*-butyldiphenylsilyl-5-deoxy-D-iditol (8). Compound **7** (189 mg, 0.258 mmol) was dissolved, under inert atmosphere, in dry THF (2 mL). PPh₃ (3 equiv., 0.774 mmol, 203 mg) was added and the mixture was cooled to 0°C. DIAD (3 equiv., 0.774 mmol, 150 μL) and diphenyl phosphoryl azide (3.2 equiv., 0.826 mmol, 180 μL) were slowly added. The mixture was stirred at rt overnight, and then the solvent was evaporated and the crude product was purified by flash chromatography (petroleum ether/ethyl acetate v/v 9/1), yielding **8** (139 mg, 71%) as colorless oil. $[\alpha]_D^{20} +12.5$ (c 0.9 CHCl₃); ¹H NMR: δ = 1.07 (s, 9 H, CH₃*t*Bu), 1.98 (s, 3 H, CH₃Ac), 3.49–3.52 [m, 1 H, C(5)-H], 3.59 [dd, 1 H, *J* = 10.3 Hz, *J* = 4.9 Hz, C(6a)-H], 3.72–3.84 [m, 4 H, C(2)-H, C(3)-H, C(4)-H, C(6b)-H], 4.23–4.26 [m, 2 H, C(1a)-H, C(1b)-H], 4.48 (d, 1 H, *J* = 11.5 Hz, CHPh), 4.55–4.58 (m, 2 H, 2CHPh), 4.65 (d, 1 H, *J* = 11.9 Hz, CHPh), 4.66 (d, 1 H, *J* = 11.1 Hz, CHPh), 4.73 (d, 1 H, *J* = 11.5 Hz, CHPh), 7.18–7.65 (m, 25 H, CHAr) ppm. ¹³C NMR: δ = 19.54 (Cq*t*Bu), 21.34 (CH₃Ac), 27.20 (CH₃*t*Bu), 63.51, 64.38 [C(1), C(6)], 63.88 [C(5)], 72.79, 75.06, 75.06 (3CH₂Ph), 75.84, 77.69, 78.83 [C(2), C(3), C(4)], 127.9–130.0 (CHAr), 132.9, 133.1 (2CqAr), 135.7, 135.8 (CHAr), 137.7, 137.8, 138.0 (3CqAr), 170.7 (C=O) ppm. IR (ν/cm⁻¹): 1739.78 (C=O), 2099.80 (N₃); MS (MALDI-TOF): *m/z* 780 [M + Na]⁺, 796 [M + K]⁺; C₄₅H₅₁N₃O₆Si (757.35): Calcd: C, 71.30; H, 6.78; N 5.54. Found: C, 71.50; H, 6.62; N, 5.39.

5-Azido-2,3,4-tri-O-benzyl-6-O-*t*-butyldiphenylsilyl-5-deoxy-D-iditol (9). Compound **8** (130 mg, 0.172 mmol) was dissolved in dry CH₃OH (2 mL) and Na (cat.) was added. The mixture was stirred at rt for 30 min and then was neutralized with amberlite IR 120-H⁺. After filtration the solvent was eliminated under reduced pressure and the residue was purified by flash chromatography (petroleum ether/ethyl acetate v/v 8/2), yielding **9** (112 mg, 91%) as colorless oil. $[\alpha]_D^{20} +9.3$ (c 1.3 CHCl₃); ¹H NMR: δ = 1.09 (s, 9 H, CH₃*t*Bu), 3.60–3.67 (m, 3 H), 3.72 [dd, 1 H, *J* = 10.4 Hz, *J* = 5.2 Hz], 3.79–3.86 (m, 3 H), 3.92 [dd, 1 H, *J* = 7.1 Hz, *J* = 4.2 Hz], 4.54 (d, 1 H, *J* = 11.4 Hz, CHPh), 4.59 (d, 1 H, *J* = 11.7 Hz, CHPh), 4.62–4.65 (m, 2 H, 2CHPh), 4.68 (d, 1 H, *J* = 11.2 Hz, CHPh), 4.75 (d, 1 H, *J* = 11.4 Hz, CHPh), 7.20–7.67 (m, 25 H, CHAr) ppm. ¹³C NMR: δ = 19.57 (Cq*t*Bu), 27.22 (CH₃*t*Bu), 61.68, 64.21 [C(1), C(6)], 63.69 [C(5)], 72.70, 75.00, 75.07 (3CH₂Ph), 77.80, 78.44, 79.40 [C(2), C(3), C(4)], 128.0–128.8 (CHAr), 130.0, 130.1 (CHAr), 133.0, 133.1 (2CqAr), 135.8–135.9 (CHAr), 137.9, 137.9, 138.0 (3CqAr) ppm. IR: 2100.0, (N₃), 3455.9 (OH); MS (MALDI-TOF): *m/z* 739 [M + Na]⁺, 755 [M + K]⁺; C₄₃H₄₉N₃O₅Si (715.34): Calcd: C, 72.14; H, 6.90; N, 5.87. Found C, 71.99; H, 6.82; N, 5.50.

5-Azido-2,3,4-tri-O-benzyl-6-O-*t*-butyldiphenylsilyl-5-deoxy-D-idose (10). Compound **9** (826 mg, 1.15 mmol) was dissolved in CH₂Cl₂ (10 mL) and Dess Martin Periodinane (1.5 equiv., 1.73 mmol, 734 mg) was added. The mixture

was stirred at rt for 30 min, then quenched with a saturated aqueous solution of NaHCO₃ (5 mL) and a 10% aqueous solution of Na₂S₂O₃ (5 mL). The mixture was stirred for 1 h, and then the two layers were separated and the organic layer was washed with a saturated solution of NaHCO₃ (2 × 10 mL) and brine (1 × 10 mL). The organic layer was dried over Na₂SO₄ and solvent was eliminated under reduced pressure. The crude product was used for the next step without further purification. $[\alpha]_{\text{D}}^{20} +20.0$ (c 1.0 CHCl₃); ¹H NMR: $\delta = 1.09$ (CH₃tBu), 3.56–3.61 [m, 1H, C(5)-H], 3.71 [dd, 1H, $J = 10.5$ Hz, $J = 7.0$ Hz, C(6a)-H], 3.77 [dd, 1H, $J = 10.5$ Hz, $J = 4.4$ Hz, C(6b)-H], 3.89 [bt, 1H, C(4)-H], 3.92–3.96 [m, 2H, C(2)-H, C(3)-H], 4.48 (s, 2H, CH₂Ph), 4.51–4.54 (m, 2H, 2CHPh), 4.62 (d, 1H, $J = 11.0$ Hz, CHPh), 4.83 (d, 1H, $J = 11.9$ Hz, CHPh), 7.26–7.67 (m, 25H, CHAr), 9.66 [s, 1H, C(1)-H] ppm. ¹³C NMR: $\delta = 19.60$ (CqtBu), 27.22 (CH₃tBu), 64.09 [C(5)], 64.15 [C(6)], 73.40, 74.42, 74.65 (3CH₂Ph), 77.12, 79.92, 80.96 [C(2), C(3), C(4)], 127.9–130.1 (CHAr), 132.9, 133.0 (2CqAr), 135.8, 135.9 (CHAr), 137.0, 137.3, 137.5 (3CqAr), 200.6 [C(1)] ppm. IR: 1728.6 (C=O), 2100.7, (N₃); MS (MALDI-TOF): m/z 737 [M + Na]⁺, 753 [M + K]⁺; C₄₃H₄₇N₃O₅Si (713.33): Calcd: C, 72.34; H, 6.64; N, 5.89. Found: C, 72.00; H, 6.72; N, 5.55.

(2R/S,3R,4S,5R,6S) Dimethyl 6-Azido-3,4,5-tris-benzyloxy-7-*t*-butyldi-phenylsilanyloxy-2-hydroxy-heptylphosphonate (11). In a flame-dried vessel, BuLi (2.30 mmol, 1.6 M in hexane, 1.44 mL) was diluted with dry THF (10 mL), and at –78°C dimethyl methyl phosphonate (2.30 mmol, 254 μ L) was slowly added. The mixture was stirred at –78°C for 30 min, and then **10** (1.15 mmol) dissolved in dry THF (8 mL) was added. The mixture was stirred at –78°C for 1 h, then quenched with NH₄Cl (1 M, 4 mL). CH₂Cl₂ (10 mL) was added, and the two layers were separated. The organic layer was washed with NH₄Cl 1M (1 × 10 mL) and H₂O (2 × 10 mL). The organic phase was dried over Na₂SO₄, and the solvent was eliminated under reduced pressure. The crude was purified by flash chromatography (petroleum ether/ethyl acetate v/v 4/6), yielding an inseparable mixture of two diastereoisomers (*R/S* 23/77, 597 mg, 62% over two steps) as colorless oil.

(2S,3R,4S,5R,6S) Dimethyl 6-Amino-3,4,5-tris-benzyloxy-7-*t*-butyldi-phenylsilanyloxy-2-hydroxy-heptylphosphonate (12). The mixture **11** (381 mg, 0.455 mmol) was dissolved in THF (8 mL). PPh₃ (2 equiv., 0.909 mmol, 239 mg) and H₂O (25 equiv., 11.38 mmol, 205 μ L) were added, and the mixture was stirred at 60°C for 18 h. The solvent was evaporated and the crude product was purified by flash chromatography (AcOEt to AcOEt/CH₃OH v/v 15/1), yielding pure **12** (190 mg) (yield 51%) as a colorless oil. $[\alpha]_{\text{D}}^{20} +2.7$ (c 1.0 CHCl₃); ¹H NMR: $\delta = 1.07$ (s, 9H, CH₃tBu), 1.91 [ddd, 1H, $J = 18.9$ Hz, $J = 15.3$ Hz, $J = 3.3$ Hz, C(1a)-H], 2.10 [ddd, 1H, $J = 16.0$ Hz,

$J = 15.3$ Hz, $J = 9.6$ Hz, C(1b)-H], 3.14–3.16 [m, 1 H, C(6)-H], 3.51 [dd, 1 H, $J = 9.6$ Hz, $J = 7.4$ Hz, C(7a)-H], 3.63–3.72 [m, 2 H, C(3)-H, C(7b)-H], 3.66 (d, 3 H, $J_{\text{H,P}} = 5.7$ Hz, OCH_3), 3.69 (d, 3 H, $J_{\text{H,P}} = 5.6$ Hz, OCH_3), 3.81 [dd, 1 H, $J = 6.1$ Hz, $J = 3.1$ Hz, C(5)-H], 4.11 [bt, 1 H, C(4)-H], 4.30–4.36 [m, 1 H, C(2)-H], 4.48 (d, 1 H, $J = 11.1$ Hz, CHPh), 4.54 (d, 1 H, $J = 11.2$ Hz, CHPh), 4.70 (s, 2 H, CH_2Ph), 4.76 (d, 1 H, $J = 11.2$ Hz, CHPh), 4.81 (d, 1 H, $J = 11.1$ Hz, CHPh), 7.20–7.65 (m, 25 H, CHAr) ppm. ^{13}C NMR: $\delta = 19.68$ (CqtBu), 27.37 (CH_3tBu), 30.19 [d, $^1J_{\text{C,P}} = 139.5$ Hz, C(1)], 52.67, 52.74 (OCH_3), 53.70 [C(6)], 66.4 [d, $^2J_{\text{C,P}} = 4.6$ Hz, C(2)], 66.98 [C(7)], 74.57, 74.78, 74.87 ($3\text{CH}_2\text{Ph}$), 78.97, 81.17, 81.31 [C(3), C(4), C(5)], 127.8–129.9 (CHAr), 133.5, 133.6 (2CqAr), 135.8 (CHAr), 138.3, 138.4, 138.5 (3CqAr) ppm. IR: 3349.6, 3361.7 (NH_2 , OH); MS (MALDI-TOF): m/z 812 [$\text{M} + \text{H}$] $^+$, 834 [$\text{M} + \text{Na}$] $^+$, 850 [$\text{M} + \text{K}$] $^+$; $\text{C}_{46}\text{H}_{58}\text{NO}_8\text{PSi}$ (811.37): calcd. C, 68.04; H, 7.20; N, 1.72. Found C, 67.97; H, 7.03; N, 1.85.

(2R,3S,4S,5R,6S) Dimethyl [3,4,5-Tris-benzyloxy-6-(*t*-butyldiphenylsilynyloxymethyl)-piperidin-2-yl]-methylenephosphonate (13).

Compound **12** (227 mg, 0.280 mmol) was dissolved under inert atmosphere in dry THF (3 mL), and then PPh_3 (2 equiv., 0.559 mmol, 147 mg) was added. The solution was cooled at 0°C , and then DIAD (2 equiv., 0.559 mmol, 108 μL) was slowly added. The mixture was stirred for 18 h at rt, and then the solvent was removed under reduced pressure and the crude was purified by flash chromatography (ethyl acetate), yielding an inseparable mixture of **13** and triphenylphosphine oxide, which was subjected to NMR analysis. ^1H NMR (400 MHz, CDCl_3 , 25°C): $\delta = 1.09$ (s, 9 H, CH_3tBu), 1.69 (ddd, 1 H, $J_{\text{H,P}} = 15.6$ Hz, $J = 14.9$ Hz, $J = 10.0$ Hz, CHP), 2.44 (bdd, 1 H, $J_{\text{H,P}} = 20.1$ Hz, $J = 14.6$ Hz, CHP), 3.11 [bt, 1 H, $J = 8.8$ Hz, C(3)-H], 3.31 [bt, 1 H, $J = 9.4$ Hz, C(2)-H], 3.38–3.41 [m, 1 H, C(6)-H], 3.62–3.67 [m, 1 H, C(4)-H], 3.68 (d, 3 H, $J_{\text{H,P}} = 10.9$ Hz, OCH_3), 3.71 (d, 3 H, $J_{\text{H,P}} = 11.1$ Hz, OCH_3), 3.75–3.79 [m, 2 H, C(5)-H, CHOH], 4.01 (bt, 1 H, CHOH), 4.34 (d, 1 H, $J = 11.5$ Hz, CHPh), 4.38 (d, 1 H, $J = 11.5$ Hz, CHPh), 4.59 (d, 1 H, $J = 11.9$ Hz, CHPh), 4.66 (d, 1 H, $J = 11.7$ Hz, CHPh), 4.82 (d, 1 H, $J = 11.7$ Hz, CHPh), 4.94 (d, 1 H, $J = 11.9$ Hz, CHPh), 7.10–7.71 (m, CHAr) ppm. ^{13}C NMR: $\delta = 19.62$ (CqtBu), 27.24 (CH_3tBu), 28.47 (d, $^1J_{\text{C,P}} = 139.6$ Hz, CH_2P), 49.46 [d, $^2J_{\text{C,P}} = 5.4$ Hz, C(2)], 52.72 (d, $^3J_{\text{C,P}} = 6.1$ Hz, OCH_3), 52.94 (d, $^3J_{\text{C,P}} = 6.1$ Hz, OCH_3), 56.40 [C(6)], 60.75 (CH_2OTBDPS), 75.60, 75.77, 75.79 ($3\text{CH}_2\text{Ph}$), 80.51 [C(5)], 83.52 [d, $^4J_{\text{C,P}} = 1.1$ Hz, C(4)], 83.77 [d, $^3J_{\text{C,P}} = 16.9$ Hz, C(3)], 127.8–128.7 (CHAr), 129.9, 130.0 (CHAr), 132.1–132.3 (CHAr), 132.2, 133.2, 133.5, 133.6 (4CqAr), 135.7, 135.8 (CHAr), 138.1, 138.3, 138.8 (3CqAr).

(2R,3S,4S,5R,6S) Dimethyl (3,4,5-Tris-benzyloxy-6-hydroxymethyl-piperidin-2-yl)-methylenephosphonate (14). Compound **13** was dissolved

in dry THF (3 mL), and then TBAF (0.560 mmol, 1 M in THF, 560 μ L) was added. The mixture was stirred at rt overnight and then quenched with buffer phosphate (3 mL). The two layers were separated, and the aqueous layer was extracted with AcOEt (3 \times 5 mL). The combined organic layers were dried over Na₂SO₄. The solvent was eliminated under reduced pressure, and the residue was purified by flash chromatography (ethyl acetate to ethyl acetate/methanol v/v 9/1), yielding **14** (90 mg, 58% over two steps) as colorless oil. $[\alpha]_D^{20} +10.1$ (*c* 1.3 CHCl₃); ¹H NMR: $\delta = 1.62$ (ddd, 1 H, *J* = 15.5 Hz, *J* = 15.3 Hz, *J* = 10.3 Hz, *CHP*), 2.30 (ddd, 1 H, *J* = 18.2 Hz, *J* = 15.1 Hz, *J* = 2.1 Hz, *CHP*), 3.02–3.10 [m, 1 H, C(2)-H], 3.14 [bt, 1 H, *J* = 8.5 Hz, C(3)-H], 3.39 [bdt, 1 H, *J* = 10.5 Hz, *J* = 5.4 Hz, *J* = 5.4 Hz, C(6)-H], 3.70 [dd, 1 H, *J* = 9.0 Hz, *J* = 8.0 Hz, C(4)-H], 3.71 (d, 3 H, *J* = 10.9 Hz, OCH₃), 3.72 (d, 3 H, *J* = 10.9 Hz, OCH₃), 3.77 [dd, 1 H, *J* = 9.0 Hz, *J* = 5.6 Hz, C(5)-H], 3.77–3.80 (m, 1 H, *CHOH*), 3.86 (bt, 1 H, *J* = 10.5 Hz, *CHOH*), 4.59 (d, 1 H, *J* = 11.2 Hz, *CHPh*), 4.66 (s, 2 H, *CH*₂Ph), 4.76 (d, 1 H, *J* = 10.9 Hz, *CHPh*), 4.90 (d, 1 H, *J* = 10.9 Hz, *CHPh*), 4.91 (d, 1 H, *J* = 11.2 Hz, *CHPh*), 7.28–7.33 (m, 15 H, *CHAr*) ppm. ¹³C NMR: $\delta = 27.27$ (d, ¹*J*_{C,P} = 140.3 Hz, *CH*₂P), 48.90 [d, ²*J*_{C,P} = 4.6 Hz, C(2)], 52.85 (d, ³*J*_{C,P} = 6.1 Hz, OCH₃), 52.97 (d, ³*J*_{C,P} = 6.1 Hz, OCH₃), 54.99 [C(6)], 58.58 (*CH*₂OH), 73.09, 75.29, 75.90 (3*CH*₂Ph), 81.31 [C(5)], 83.19 [d, ³*J*_{C,P} = 16.1 Hz, C(3)], 83.61 [d, ⁴*J*_{C,P} = 2.3 Hz, C(4)], 127.9–128.7 (*CHAr*), 138.1, 138.3, 138.7 (3*CqAr*) ppm. MS (MALDI-TOF): *m/z* 556 [M + H]⁺, 578 [M + Na]⁺, 594 [M + K]⁺; C₃₀H₃₈NO₇P (555.24): Calcd: C, 64.85; H, 6.89; N, 2.52. Found: C, 64.69; H, 6.77; N, 2.61.

1-Deoxy-L-idonojirimycin-1-methylenphosphonate (1). Compound **14** (50 mg, 0.09 mmol) was dissolved in MeOH (3 mL); a catalytic amount of Pd(OH)₂ and acetic acid (1 mL) were added and then the reaction mixture was stirred under H₂ overnight. The catalyst was filtered through a pad of Celite (eluting with MeOH) and then the solvent was evaporated under reduced pressure to afford pure compound **1** (26 mg, 98% yield) as an amorphous solid. $[\alpha]_D^{20} -22.4$ (*c* 1.0 MeOH); ¹H NMR (D₂O): $\delta = 2.32$ (ddd, 1 H, *J* = 19.4 Hz, *J* = 16.3 Hz, *J* = 6.6 Hz, *CHP*), 2.61 (ddd, 1 H, *J* = 20.4 Hz, *J* = 16.3 Hz, *J* = 5.8 Hz, *CHP*), 3.16–3.28 [m, 2 H, C(2)-H, C(6)-H], 3.44–3.51 [m, 2 H, C(3)-H, C(4)-H], 3.66 (d, 3 H, *J* = 11.1 Hz, OCH₃), 3.67 (d, 3 H, *J* = 11.1 Hz, OCH₃), 3.74–3.79 (m, 2 H, *CH*₂OH), 3.84–3.90 [m, 1 H, C(5)-H] ppm. ¹³C NMR (D₂O): $\delta = 25.10$ (d, ¹*J*_{C,P} = 141.8 Hz, *CH*₂P), 54.33 [d, ²*J*_{C,P} = 6.0 Hz, C(2)], 56.11 (d, ³*J*_{C,P} = 6.9 Hz, POCH₃), 56.18 (d, ³*J*_{C,P} = 6.9 Hz, POCH₃), 56.53 [C(6)], 58.67 (*CH*₂OH), 70.88 [C(5)], 73.09 [d, ³*J*_{C,P} = 11.3 Hz, C(3)], 76.20 [C(4)] ppm. ³¹P NMR (D₂O): $\delta = 39.11$ ppm; MS (MALDI-TOF): *m/z* 286 [M + H]⁺; C₉H₂₀NO₇P (285.10): calcd: C, 37.90; H, 7.07; N, 4.91. Found: C, 38.01; H, 6.84; N, 5.03.

REFERENCES

- [1] (a) Lillelundt, V.H.; Jensen, H.H.; Liang, X.; Bols, M. Recent developments of transition-state analogue glycosidase inhibitors of non-natural product origin. *Chem. Rev.* **2002**, *102*, 515–554; (b) Asano, N.; Nash, R.J.; Molyneux, R.J.; Fleet, G.W.J. Sugar-mimic glycosidase inhibitors: natural occurrence, biological activity and prospects for therapeutic application. *Tetrahedron: Asymm.* **2000**, *11*, 1645–1680; (c) Asano, N. Alkaloidal sugar mimetics: biological activities and therapeutic applications. *J. Enzyme Inhib.* **2000**, *15*, 215–234; (d) Zechel, D.L.; Withers, S.G. Glycosidase mechanisms: anatomy of a finely tuned catalyst. *Acc. Chem. Res.* **2000**, *33*, 11–18; (e) Heightman, T.D.; Vasella, A.T. Recent insights into inhibition, structure, and mechanism of configuration-retaining glycosidases. *Angew. Chem. Int. Ed.* **1999**, *38*, 750–770; (f) Sears, P.; Wong, C.-H. Carbohydrate mimetics: a new strategy for tackling the problem of carbohydrate-mediated biological recognition. *Angew. Chem. Int. Ed.* **1999**, *38*, 2300–2324; (g) Stütz, A.E. *Iminosugars as Glycosidase Inhibitors: Nojirimycin and Beyond*; Wiley-VCH: Weinheim, Germany, 1999; (h) Asano, N.; Nishida, M.; Kato, A.; Kizu, H.; Matsui, K.; Shimada, Y.; Itoh, T.; Baba, M.; Watson, A.A.; Nash, R.J.; Lilley, P.M.de Q.; Watkin, D.J.; Fleet, G.W.J. Homonojirimycin isomers and n-alkylated homonojirimycins: structural and conformational basis of inhibition of glycosidases. *J. Med. Chem.* **1998**, *41*, 2565–2571; (i) Ganem, B. Inhibitors of carbohydrate-processing enzymes: design and synthesis of sugar-shaped heterocycles. *Acc. Chem. Res.* **1996**, *29*, 340–347.
- [2] (a) Ratner, L.; vander Heyden, N.; Dederá, D. Inhibition of HIV and SIV infectivity by blockade of alpha-glucosidase activity. *Virology* **1991**, *181*, 180–192; (b) Rudd, P.M.; Elliott, T.; Cresswell, P.; Wilson, I.A.; Dwek, R.A. Glycosylation and the immune system. *Science* **2001**, *291*, 2370–2376; (c) Chery, F.; Cronin, L.; O'Brien, J.L.; Murphy, P.V. Synthesis of peptidomimetics based on iminosugar and β -d-glucopyranoside scaffolds and inhibitor of HIV-protease. *Tetrahedron* **2004**, *60*, 6597–6608; (d) Greimel, P.; Spreitz, J.; Stutz, A.E.; Wrodnigg, T.M. Iminosugars and relatives as antiviral and potential anti-infective agents. *Curr. Topics Med. Chem.* **2003**, *3*, 513–523; (e) Alper, J. Searching for medicine's sweet spot. *Science* **2001**, *291*, 2338–2343; (f) Pavlović, D.; Neville, D.C.A.; Argaud, O.; Blumberg, B.; Dwek, R.A.; Fischer, W.B.; Zitzmann, N. The hepatitis C virus p7 protein forms an ion channel that is inhibited by long-alkyl-chain iminosugar derivatives. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 6104–6108; (g) Durantel, D.; Carrouee-Durantel, S.; Branza-Nichita, N.; Dwek, R.A.; Zitzmann, N. Effects of interferon, ribavirin, and iminosugar derivatives on cells persistently infected with noncytotoxic bovine viral diarrhoea virus. *Antimicrob. Agents Chemother.* **2004**, *48*, 497–504; (h) Wu, S.-F.; Lee, C.-J.; Liao, C.-L.; Dwek, R.A.; Zitzmann, N.; Lin, Y.-L. Antiviral effects of an iminosugar derivative on flavivirus infections. *J. Virol.* **2002**, *76*, 3596–3604.
- [3] (a) Paulsen, H.; Brockhausen, I. From imino sugars to cancer glycoproteins. *Glycoconj. J.* **2001**, *18*, 867–870; (b) Goss, P.E.; Baker, M.A.; Carver, J.P.; Dennis, J.W. Inhibitors of carbohydrate processing: a new class of anticancer agents. *Clin. Cancer Res.* **1995**, *1*, 935–944.
- [4] (a) Anzeveno, P.B.; Creemer, L.J.; Daniel, J.K.; King, C.H.R.; Liu, P.S. A facile, practical synthesis of 2,6-dideoxy-2,6-imino-7-O- β -D-glucopyranosyl-D-glycero-L-gulo-heptitol (MDL 25,637). *J. Org. Chem.* **1989**, *54*, 2539–2542; (b) Balfour, J.A.; McTavish, D. Acarbose. An update of its pharmacology and therapeutic use in diabetes mellitus. *Drugs* **1993**, *46*, 1025–54.

- [5] (a) Cren, S.; Gurcha, S.S.; Blake, A.J.; Besra, G.S.; Thomas, N.R. Synthesis and biological evaluation of new inhibitors of UDP-Gal₄ transferase—a key enzyme in *M. tuberculosis* cell wall biosynthesis. *Org. Biomol. Chem.* **2004**, *2*, 2418–2420; (b) Wrodnigg, T.M.; Sprenger, F.K. Bioactive carbohydrates and recently discovered analogues as chemotherapeutics. *Mini-Rev. Med. Chem.* **2004**, *4*, 437–459.
- [6] (a) Butters, T.D.; van den Broek, L.A.G.M.; Fleet, G.W.J.; Krulle, T.M.; Wormald, M.R.; Dwek, R.A.; Platt, F.M. Molecular requirements of imino sugars for the selective control of N-linked glycosylation and glycosphingolipid biosynthesis. *Tetrahedron: Asymm.* **2000**, *11*, 113–124; (b) Butters, T.D.; Dwek, R.A.; Platt, F.M. Inhibition of glycosphingolipid biosynthesis: application to lysosomal storage disorders. *Chem. Rev.* **2000**, *100*, 4683–4696; (c) Cox, T.M.; Aerts, J.M.F.G.; Andria, G.; Beck, M.; Belmatoug, N.; Bembi, B.; Chertkoff, R.; Vom Dahl, S.; Elstein, D.; Erikson, A.; Giralt, M.; Heitner, R.; Hollak, C.; Hrebicek, M.; Lewis, S.; Mehta, A.; Pastores, G.M.; Rolfs, A.; Sa Miranda, M.C.; Zimran, A. The role of the iminosugar N-butyldeoxynojirimycin (miglustat) in the management of type I (non-neuronopathic) Gaucher disease: a position statement. *J. Inherit. Metab. Dis.* **2003**, *26*, 513–526.
- [7] (a) Bouix, C.; Bissret, P.; Eustache, J. Stereoselective synthesis of Arabinose-derived phosphonates. *Tetrahedron Lett.* **1998**, *39*, 825–828; (b) Bosco, M.; Bissret, P.; Bouix, C.; Eustache, J. A new concise synthesis of nectrisine and its facile conversion to phosphonoazasugars. *Tetrahedron Lett.* **2001**, *42*, 7949–7952; (c) Godin, G.; Compain, P.; Masson, G.; Martin, O.R. A general strategy for the practical synthesis of nojirimycin C-glycosides and analogues. Extension to the first reported example of an iminosugar 1-phosphonate. *J. Org. Chem.* **2002**, *67*, 6960–6970.
- [8] Ramirez, F.; Mandal, S.B.; Marecek, J.F. Synthesis of phosphatidyl-6-D-glucose and attempted synthesis of phosphatidyl-1-D-glucose. *J. Org. Chem.* **1983**, *48*, 2008–2013.